

Studies in Fungi 3(1): 115–120 (2018) www.studiesinfungi.org ISSN 2465-4973 **Article**

Doi 10.5943/sif/3/1/13

Copyright © Institute of Animal Science, Chinese Academy of Agricultural Sciences

Kamalomyces polyseptatus sp. nov. from an unidentified bamboo twig in Andaman Islands, India

Niranjan M and Sarma VV*

Department of Biotechnology, Pondicherry University, Kalapet, Pondicherry-605014, India.

Niranjan M, Sarma VV 2018 – *Kamalomyces polyseptatus* sp. nov. from an unidentified bamboo twig in Andaman Islands, India. Studies in Fungi 3(1), 115–120, Doi 10.5943/sif/3/1/13

Abstract

The examination of decaying twig samples fallen on the forest floor in the Andaman Islands, India yielded a new fungal species in the genus *Kamalomyces* (Tubeufiaceae). The new species, *Kamalomyces polyseptatus* is described in this paper supported by photomicrographs. The novel species is characterized by superficial, scattered, globose to sub-globose, dark brown to black ascomata covered by dense black mycelium, clavate asci, vermiform, elongate, transverse septate, hyaline ascospores. The new species *K. polyseptatus* is easily distinguishable from other species of the genus by the presence of higher number of septa in the ascospores. A synopsis of salient features of different species of *Kamalomyces* is provided along with a dichotomous key to the known species of *Kamalomyces*.

Key words – Bamboo twigs – Dothideomycetes – Taxonomy – *Tubeufiaceae*

Introduction

The order Tubeufiales was introduced by Boonmee et al. (2014) based on morphology and molecular phylogeny to accommodate the family Tubeufiaceae. This family was established by Barr (1979) based on the type genus Tubeufia to accommodate bitunicate ascomycetes, with superficial ascomata surrounded by hyphae or setae, growing as saprobes on decaying wood and producing phragmosporous ascospores (Barr 1979, Phookamsak et al. 2017). Currently 25 genera have been accepted in this family (Wijayawardene et al. 2018) including Acanthohelicospora, Acanthophiobolus, Acanthostigma, Acanthostigmina, Aquaphila, Berkleasmium, Bifrontia, Boerlagiomyces, Chaetosphaerulina, Chlamydotubeufia, Dictyospora, Helicangiospora, Helicoma, Helicomyces, Helicosporium, Kamalomyces, Muripulchra, Neoacanthostigma, Neohelicomyces, Neotubeufia, Podonectria, Tamhinispora, Thaxteriella, Thaxteriellopsis and Tubeufia. Most of the genera have uniloculate, superficial, pigmented ascomata (Boonmee et al. 2014). Earlier Tubeufiaceae was placed in Pleosporales (Barr 1979) but was later transferred to Dothideales (Eriksson & Winka 1998). Subsequently, based on phylogenetic analysis of 28s rDNA, Kodsueb et al. (2006) placed the family in Pleosporales once again. Boonmee et al. (2011) revisited the family and examined the type and rDNA phylogeny, which showed that it has close relationship with Botryosphaeriales. Finally, Boonmee et al. (2014) raised a new order Tubeufiales to accommodate Tubeufiaceae and added three new genera Acanthohelicospora, Helicangiospora and *Neoacanthostigma* with sexual and asexual morphs connections.

Kamalomyces was introduced by Verma et al. (2008) with Kamalomyces indicus as the type species recorded on bamboo. Recently three new species were added in Kamalomyces: K.

mahabaleshwarensis, K. thailandicus and K. bambusicola (Dubey & Neelima 2013, Phookamsak et al. 2017). Kamalomyces is distinct from the other genera of Tubeufiaceae (Phookamsak et al. 2017) in the occurrence of sub-globose to lemoniform ascomata with solitary-clusters surrounded by hyphae, broadly cylindrical to clavate asci and hyaline, vermiform, septate ascospores. Kamalomyces indicus and K. thailandicus have vermiform spores (Phookamsek et al. 2017) while the remaining two species: K. bambusicola and K. mahabaleshwarensis have non-vermiform ascospores (Phookamsak et al. 2017). Boonmee et al. (2014) proposed a key to the genera of Tubeufiaceae.

We are investigating the diversity of wood degrading, saprophytic, filamentous ascomycetous fungi from Andaman Islands, India. In our recent collection we found a specimen that has characters similar to the genus *Kamalomyces*. Upon closer examination it has been found that the new collection does not fit in any of the existing species in the genus *Kamalomyces* and hence it is reported as a new species in this paper.

Material and methods

Dead and decaying twig samples fallen on the forest floor in the reserved forests of North Andaman Islands, India were collected in January 2017 and transferred into zip lock plastic bags, air dried overnight, and packed into new plastic bags for shipment to the laboratory for further processing. Before undertaking the microscopic examination, the twigs were placed individually into plastic bread boxes lined with sterile tissue paper, rehydrated by sprinkling sterile water and incubated for one week to three months. The samples are then examined under a Stereo Zoom microscope (Optika SZM-LED, Italy) to locate the fungal fruiting structures. Hand sections were taken wherever necessary. The fruit bodies were cut with a razor and the spore constituents were transferred to a micro slide mounted with stains like Lacto phenol, Lacto phenol cotton blue, Lougal's reagent. These slides were then examined under the Nikon ECLIPSE TiU upright microscope with DIC objectives fitted with Nikon DS-Fi2 digital camera, Japan to take photomicrographs. Measurements were taken with Nikon NIS-Elements-Imaging Software version 4.4 program. Photo plates were prepared and edited with Microsoft power point, and Adobe Photoshop version 7.0. Morphological identification was carried out by referring to various individual publications (Verma et al. 2008, Dubey & Neelima 2013, Phookamsak et al. 2017, Kirk et al. 2008). The herbarium materials were deposited at Ajrekar Mycological Herbarium (AMH), Agharkar Research Institute (ARI), Pune, India.

Kamalomyces polyseptatus M. Niranjan and V.V. Sarma sp. nov.

Mycobank: MB824916

Etymology – With reference to the multiple septate ascospores

Saprobic on an un-identified bamboo twig. Sexual morph - Colonies 8-20×10-15 cm, superficial, surrounded by dense black, hyphal mat with swollen brown cells, apically raised, loosely connected, brown, septate, thick-walled, hyphae 4.6 µm wide. Ascomata 230–270 × 200– 260 µm, perithecial, globose to sub-globose, basal dark brown, lateral and apical parts brown, immersed in the hyphae, coriaceous, scattered. Peridium 22.1 µm wider, consists two layers, outer rough, brown layer of textura angularis cells with munk pores and an inner layer of hyaline textura angularis cells. Hamathecium: Pseudoparaphyses 1.7µm wide, filamentous, anastomosing, longer than asci, interspersed within gelatinous matrix. Asci 154–224 (247.7) \times 25.2–33.5 (\bar{X} = 186.4 \times 29.5) µm (n=25), bitunicate, fissitunicate, broad clavate to clavate, with thick ocular chamber covered by smooth wall layer, pedicellate, pedicel long when young, becomes short at maturity. Ascospores 83–99.5 (104.2) \times 6.8–8.4 (8.8) ($\overline{X} = 91.3 \times 7.7$) µm (n=25), multi-seriate, vermiform, hyaline to pale brown, fusiform-clavate, apically obtuse, basally with narrow ends, mostly curved to straight, smooth-walled, two layered, phragmosporous, distoseptate, pleomorphic spores with 3 recognizable types, in first type apical 4-9 cells are wider, second type only 6th cell is wider, third type mostly 5th cell enlarged along with 11 and 12 or 16 and 18th also enlarged. Swollen cells are 8–9.9 μ m ($\bar{X} = 8.9$) (n=25) wide. Asexual morph–Undetermined.

Known distribution – India

Material examined – India, Andaman and Nicobar Islands, North Andaman, Khalighat, (13°13'43.9"N 92°56'35.3"E). Isolated from an unidentified bamboo twig (Holotype AMH 9968), 6 January, 2017, Niranjan M & Sarma VV.

Notes – Kamalomyces polyseptatus is different from K. indicus in having hyaline ascospores with supramedian swollen cells (Figs 1, 2). Spores are similar to K. bambusicola excepting the presence of a supramedian swollen cell in K. polyseptatus. Kamalomyces polyseptatus has more septa than any other existing species (Table 1). Kamalomyces polyseptatus having the 5th cell swollen is similar to K. bambusicola, but differs from the latter in having two more cells that are wider such as 11th and 12th or 16 and 18th cells. Kamalomyces polyseptatus is different from K. thailandicus in having hyphal subiculum and more than one swollen cell and from K. mahabaleshwarensis in having larger ascospores and more septa (Table 1).

Discussion

The family Tubeufiaceae was established by Barr (1979) who placed it in Pleosporales and included 5 genera. Recently Boonmee et al. (2011) carried out a revision on this family and included 20 genera based on morphological and molecular criteria. Many of the genera of this family have uniloculate, superficial, pigmented ascomata e.g. pale brown, brown, and dark brown to black; multi-celled, hyaline ascospores, and produce, helicosporous asexual morph. Most species in this family are saprobic on terrestrial woody substrata (Boonmee et al. 2011, 2014). Based on their distinct morphology as well as combined LSU, SSU and TEF1 sequence analyses Boonmee et al. (2014) established a new order Tubeufiales to accommodate Tubeufiaceae. A dichotomous key has been provided by Boonmee et al. (2014) to delineate different genera in Tubeufiaceae. Our present collection fits very well into the genus Kamalomyces based on the morphological characters. Upon closer examination it was found that it does not fit into any of the existing species of the genus Kamalomyces and hence a new species Kamalomyces polyseptatus has been proposed to be accommodated in this genus. The differences among different species of Kamalomyces including K. polyseptatus have already been discussed in the 'notes' section above in addition to enlisting them in Table 1. There were slight overlapping characters found between K. polyseptatus and K. indicus in ascospore dimensions. However, when 'average length and width ratio' of ascospores was taken into consideration there were sharp differences found between these two species which are shown in Table 1 and also in the dichotomous key provided at the end. These were 99 x 9 μm in K. indicus vs. 91.3 x 7.7 μm in K. polyseptatus. This difference, in addition to the higher number of septa found in K. polyseptatus clearly distinguish K. polyseptatus from K. indicus. Hence a new species K. polyseptatus has been proposed to be accommodated in *Kamalomyces*, which is justified based on the above mentioned characters.

Table 1 Synopsis of important characteristics of different species belonging to *Kamalomyces*.

	Fungi	Ascomata	Asci	Ascospores	Septa no.
1	Kamalomyces indicus	(169–) 216.5–253	(140–) 157–214	(92–) 95.5 – 104.5	38–42
		× (184–) 225.5 –	$(-223) \times (27.5) 29$	$(-107) \times 8 - 10.5 \mu m$	
		295.5 (-331) μm	36 (-38) μm	$(\bar{X} = 99 \times 9 \mu m)$	
2	Kamalomyces	320-460 × 360-	170–290 × 24–34	$55-86 \times 8-12 \ \mu m$	20
	mahabaleshwarensis	450 μm	μm		
3	Kamalomyces	250 - 300 × 220-	(150–) 180–190	(57–) 75 – 90 (–98) × 4–	(20-) 27-
	bambusicola	300 μm	$(-215) \times (21-) 23-$	$7 (\bar{X} = 83.8 \times 5.5) \mu m$	30
			28 (-30) μm		
4	Kamalomyces	180 - 250 × 250-	(174–) 185–220	(100–) 120 – 135 (–155)	(19–) 33–
	thailandicus	280 μm	$(-243) \times (32-) 35-$	\times 8–10 (\bar{X} = 129 \times 8.8	36
			42 μm	μm)	
5	Kamalomyces	230–270 × 200 –	154-224 (247.7) ×	83 – 99.5 (104.2) × 6.8–	42–46
	polyseptatus	260 μm	25.2–33.5 μm	$8.4 (8.8) (\overline{X} = 91.3 \times$	
				7.7) µm	

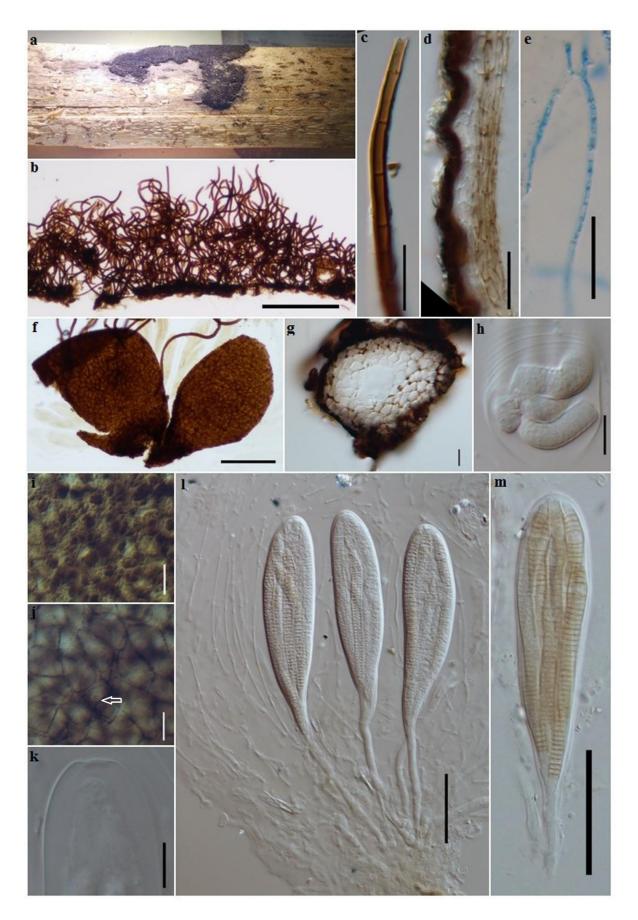


Fig. 1 – *Kamalomyces polyseptatus* (AMH-9968 Holotype). a Colony on host. b Vertical section of subiculum. c Hypha d Peridium. e Pseudoparaphyses. f, g Ascomata. h Transverse section of asci. i Peridium *textura angularis*. j peridium layer with munk pores. k Ascus with apical chamber. l-m Asci. Bars – b = 200 μ m, f = 100 μ m, l, m = 50 μ m, c-e, h, k = 20 μ m, g, i, j = 10 μ m.

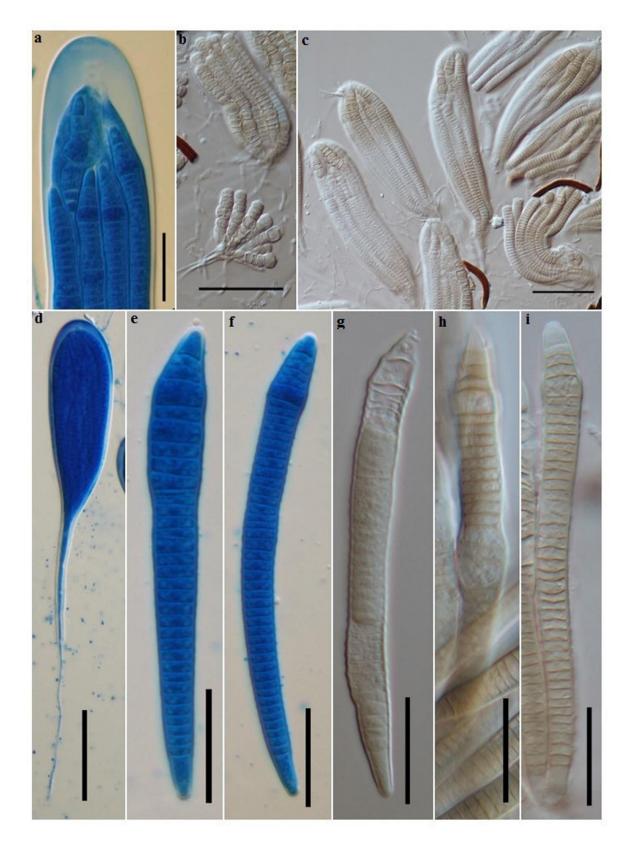


Fig. 2 – *Kamalomyces polyseptatus* a-d Asci. e-i Ascospores. (e=type 1, f,g type 2, a,g,h type 3) Bars – b–d = $50\mu m$, a, e–i = $20 \mu m$.

Dichotomous key to different species of Kamalomyces				
1. Ascospores with less than 30 septa				
1' Ascospores with more than 30 septa				
2. Ascospores dia more than 8 µm	K. mahabaleshwarensis			
2' Ascospores dia more than 8 µm				

- 4. Ascospores with less than 36 septa, average length of ascospores is 129µmK. thailandicus

Acknowledgements

We express our sincere thanks to the Science and Engineering Research Board (SERB), Ministry of Science and Technology, Govt. of India for funding the project (SERB/SB/SO/PS/18/2014 dt.19.5.2015). We are also thankful to SERB-DST, Government of India for the award of the fellowship to M. Niranjan to carry out this work. We also thank the Department of Biotechnology, Pondicherry University for providing the facilities.

References

- Barr ME. 1979 A classification of Loculoascomycetes. Mycologia. 1, 935–957.
- Boonmee S, Rossman AY, Liu JK, Li et al. 2014 Tubeufiales, ord. nov., integrating sexual and asexual generic names. Fungal Diversity 68, 239–298.
- Boonmee S, Zhang Y, Chomnunti P, Chukeatirote E et al. 2011 Revision of lignicolous Tubeufiaceae based on morphological re-examination and phylogenetic analysis. Fungal Diversity 51, 63–102.
- Dubey R, Neelima AM. 2013 *Kamalomyces mahabaleshwarensis* sp. nov. (Tubeufiaceae) from the Western Ghats, India. Mycosphere. 4, 760–763.
- Eriksson OE, Winka K. 1998 Families and higher taxa of Ascomycota. Myconet 1, 17–24.
- Kirk PM, Cannon PF, Minter DW, Stalpers JA. 2008 Ainsworth & Bisby's dictionary of the fungi, 10th edn. CABI, Wallingford.
- Kodsueb R, Jeewon R, Vijaykrishna D, McKenzie EHC et al. 2006 Systematic revision of Tubeufiaceae based on morphological and molecular data. Fungal Diversity 21, 105–130.
- Phookamsak R, Lu YZ, Hyde KD, Jeewon R et al. 2017 –Phylogenetic characterization of two novel *Kamalomyces* species in Tubeufiaceae (Tubeufiales). Mycological Progress 17, 647–660.
- Verma RK, Sharma N, Soni KK, Jamaluddin. 2008 Forest fungi of Central India. International Book Distributing Co., Lucknow, India. 418 pp.
- Wijayawardene NN, Hyde KD, Lumbsch HT, Liu JK et al. 2018 Outline of Ascomycota 2017. Fungal Diversity. 88, 167–263.